



Il biofilm microbico nelle infezioni croniche

Enea G. Di Domenico

Microbiology and Virology
San Gallicano Institute
I.F.O. Istituti Fisioterapici Ospitalieri

Hot Spots for Germs



Smartphones: Think of All the Places They've Been (and What We Touch Before Using Them)



The smartphone screen was found at reach 254.9 CFU/cm² (10 times more germs than found on a standard toilet seat)

94.5 % of the phones were contaminated with some kind of bacteria, many of which were resistant to multiple antibiotics. *S. aureus* strains isolated from mobile phones of 52% and those strains isolated from hands of 37.7% were methicillin resistant.

Your Office, Along With Other Communal Surfaces, Are Likely Teeming With Germs



4800 swabs from surfaces in office buildings, bathroom door handles to photocopier buttons to communal microwaves, refrigerators, and coffee pots

The average desktop harbours 20,961 germs per square inch and that's in addition to 3,295 on the keyboard and 1,676 on a mouse and a staggering 25,127 on the phone.

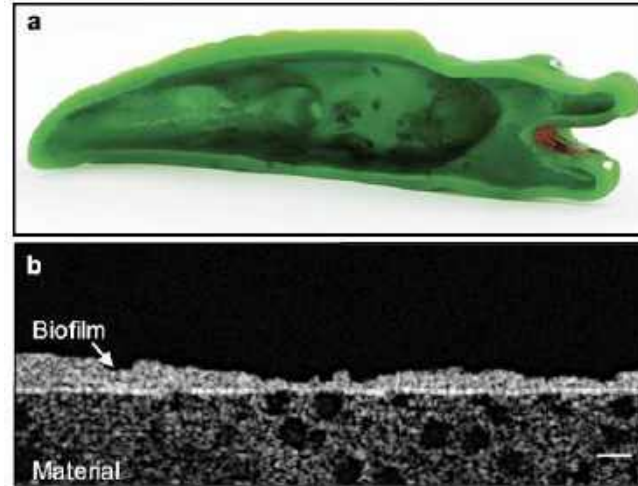
The typical office desk harbours more than 10 million bacteria (400 times more germs than found on a standard toilet seat)

Ugly ducklings—the dark side of plastic materials

The average total bacterial number was 9.5×10^6 CFU/cm².
(400 000 times more germs than found on a standard toilet seat)

Bacterial community compositions showed the presence of many rare taxa in real bath toys.

Fungi were identified in 58% of all real bath toys.



Do you really want me to shake your hand ?

There are as many as 1,000 resident bacteria per cm² on a hand

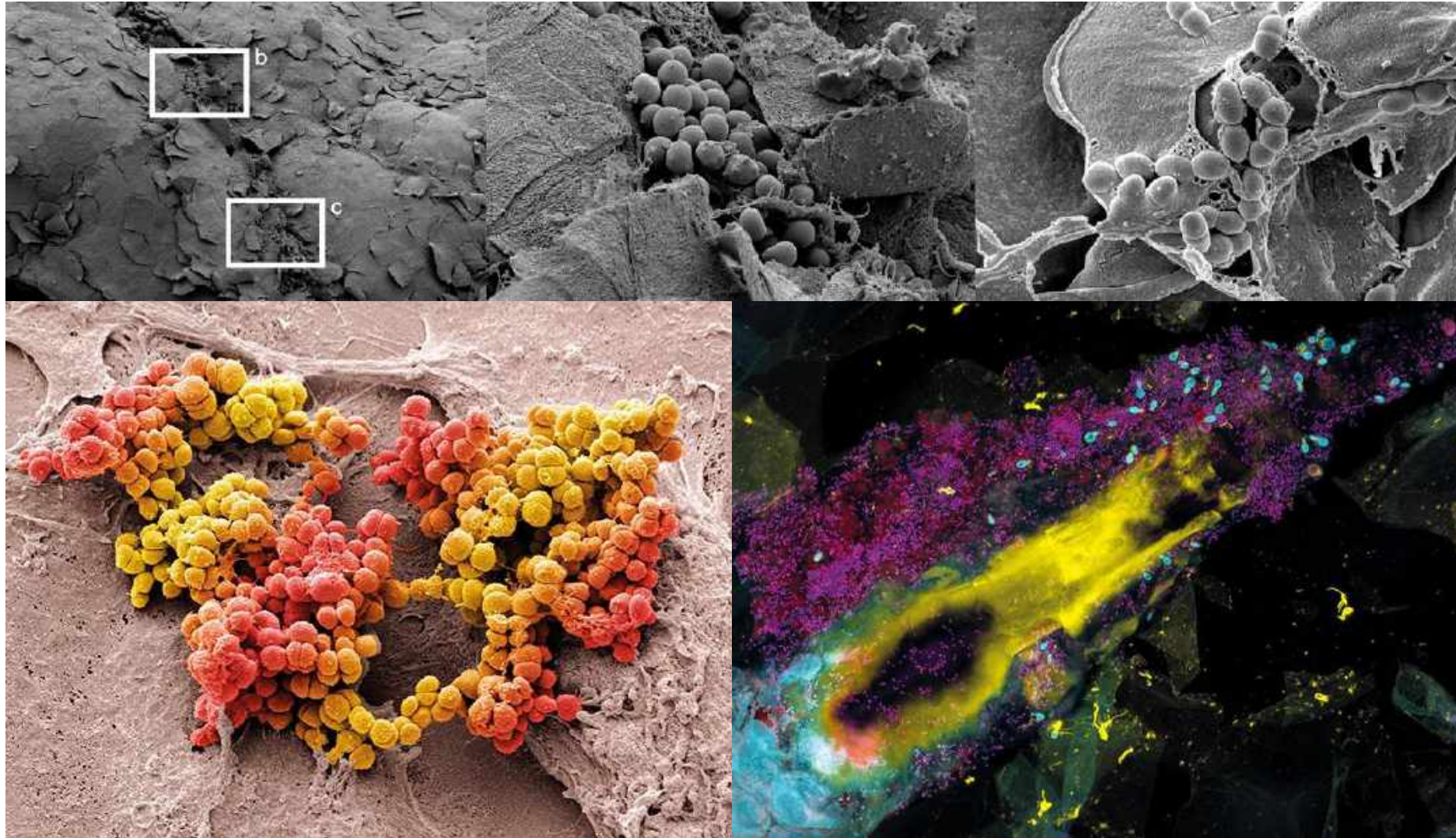
The average hand harboured 150 species of bacteria



The left and right hands of the same individual shared only about 17 per cent of the same bacteria types

The palms of women had a more diverse bacterial composition than those of men

How do they live?



The Biofilm theory

How Bacteria Stick

In nature (but not in laboratory cultures) bacteria are covered by a "glycocalyx" of fibers that adhere to surfaces and to other cells. Adhesion might be prevented by a new kind of antibiotic

by J. W. Costerton, G. G. Geesey and K.-J. Cheng

Bacteria stick tenaciously and often with exquisite specificity, to surfaces ranging from the human tooth or lung and the intestine of a cow to a rock submerged in a fast-moving stream. They do so by means of a mass of tangled fibers of polysaccharides, or branching sugar molecules, that extend from the bacterial surface and form a fuzzy "glycocalyx" surrounding an individual cell or a colony of cells. The adhesion mediated by the glycocalyx determines particular locations of bacteria in most natural environments;

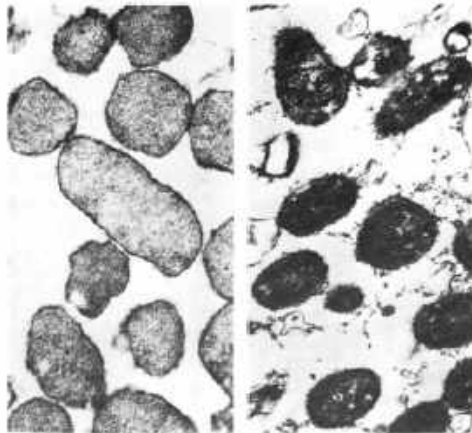
more specifically, it is a major determinant in the initiation and progression of bacterial diseases ranging from dental caries to pneumonia.

These major—and, with the benefit of hindsight, rather obvious—facts about the bacterial cell surface have become known only within the past decade. Ironically the main reason for the late discovery of the bacterial glycocalyx and its functions was the long reliance by microbiologists on an otherwise eminently effective investigative system: the pure laboratory culture of an individual

bacterial strain. To generate and maintain a glycocalyx a bacterial cell must expend energy, and in the protected environment of a pure culture the glycocalyx is a metabolically expensive luxury conferring no selective advantage; cells that fabricate these diabolite coatings are usually eliminated from pure cultures by uncoated mutants that can devote more of their energy budget to proliferation. Microbiologists have largely studied such naked mutants.

In a competitive natural environment populated by several kinds of bacteria, on the other hand, selection favors cells that are protected, and enabled to adhere to a desirable surface, by a glycocalyx. It was only in 1969 that Ivan L. Roth of the University of Georgia demonstrated carbohydrate fibers surrounding bacteria in an aquatic system and Ian W. Sutherland of the University of Edinburgh Medical School characterized the surface polysaccharides of bacteria taken from natural environments, thus drawing attention to the universality of what we now know as the glycocalyx. Since then studies in our laboratories at the University of Calgary and at the Agriculture Canada Research Station at Lethbridge in Alberta, and in a number of other laboratories, have made it clear that the glycocalyx is essential to the biological success of most bacteria in most of the varied natural environments in which they are observed.

The polysaccharide-coated surface is not a peculiarity of the bacterial cell. The more rigid polysaccharide cell wall of higher plants was among the first microscopic structures described by Robert Hooke in 1665. The analogous surface of animal cells—a glycocalyx like that of bacteria—was described only in 1971, by Vincent T. Marchesi and his colleagues at the National Institute of Arthritis, Metabolism, and Digestive Diseases. They isolated and identified glycoproteins arrayed in the membrane of animal cells and showed that the polysaccharide fibers they bear extend outward from the membrane to form



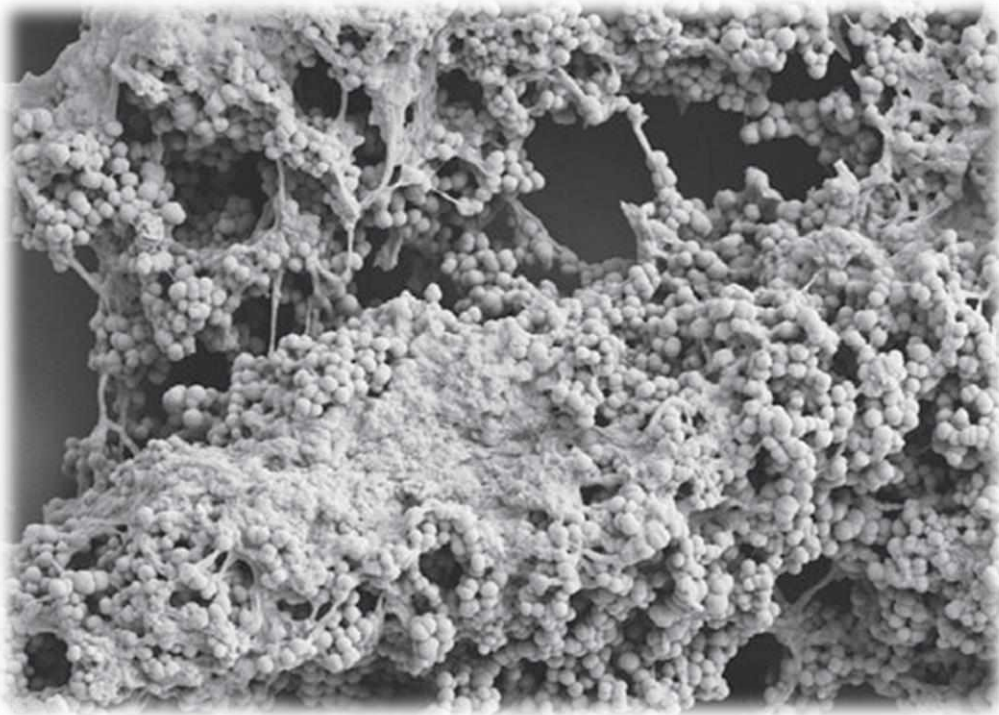
NAKED BACTERIA (left) are from a typical pure laboratory culture of *Escherichia coli*; the glycocalyx-coated bacteria (right) are from diseased cells from a infected human bladder. In both preparations the cells were stained with ruthenium red, which is taken up by a polysaccharide glycocalyx fibers that are present. The bacterial glycocalyx was ignored until recently because the familiar pure laboratory strains do not need it and therefore do not fabricate it.



In 1978 Bill Costerton warned that chronic infections in patients with indwelling medical devices were caused by biofilm bacteria and that bacteria within biofilms resist antibiotic therapies and immune host defenses

What is a Biofilm?

A biofilm is made up of colonies of one or more type of microorganisms that form together under a sticky glue-like substance and attach to all kinds of surfaces. Microorganisms such as bacteria and fungi, commonly make up biofilm colonies.



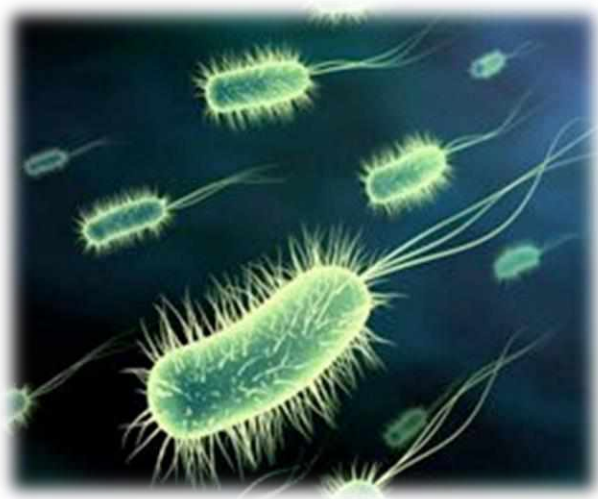
Biofilm cells are frequently embedded within a self-produced matrix of extracellular polymeric substance (EPS).

Biofilm EPS, is a polymeric jumble of DNA, proteins and polysaccharides.

Biofilms: The social life of microorganisms

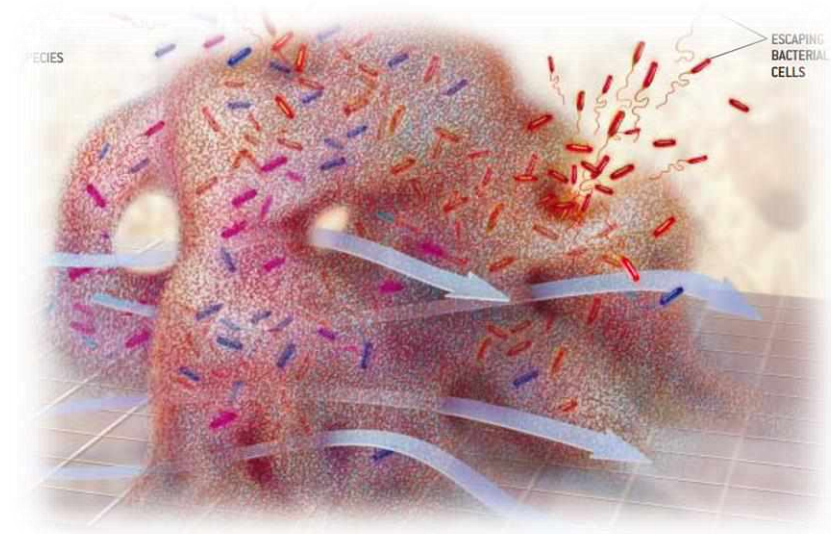
Biofilms formation provides significant benefits in terms of host colonization, defence against competitors and adaptation to changing environments

Planktonic bacteria



Extracellular matrix

Bacterial biofilm

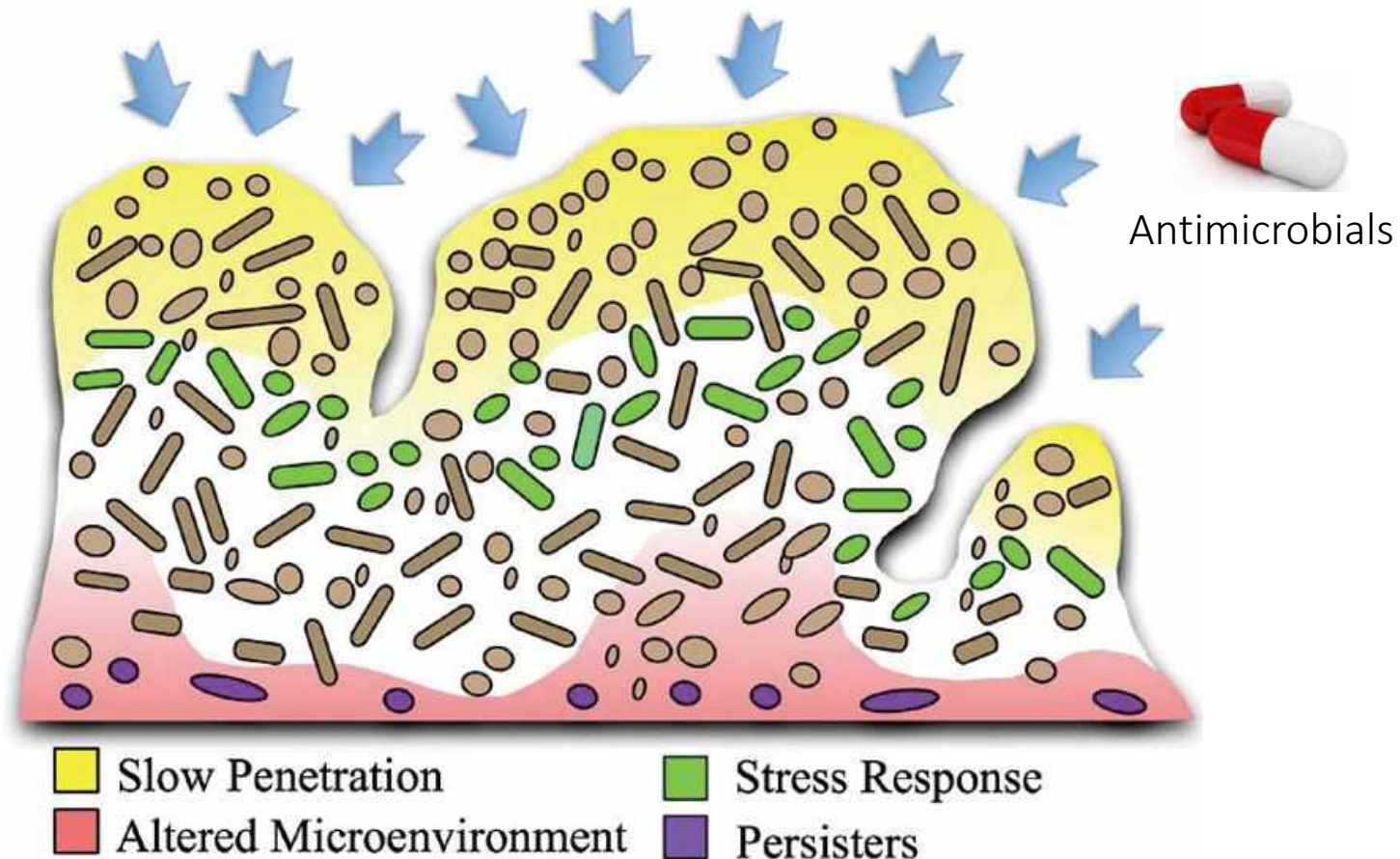


Growth rates

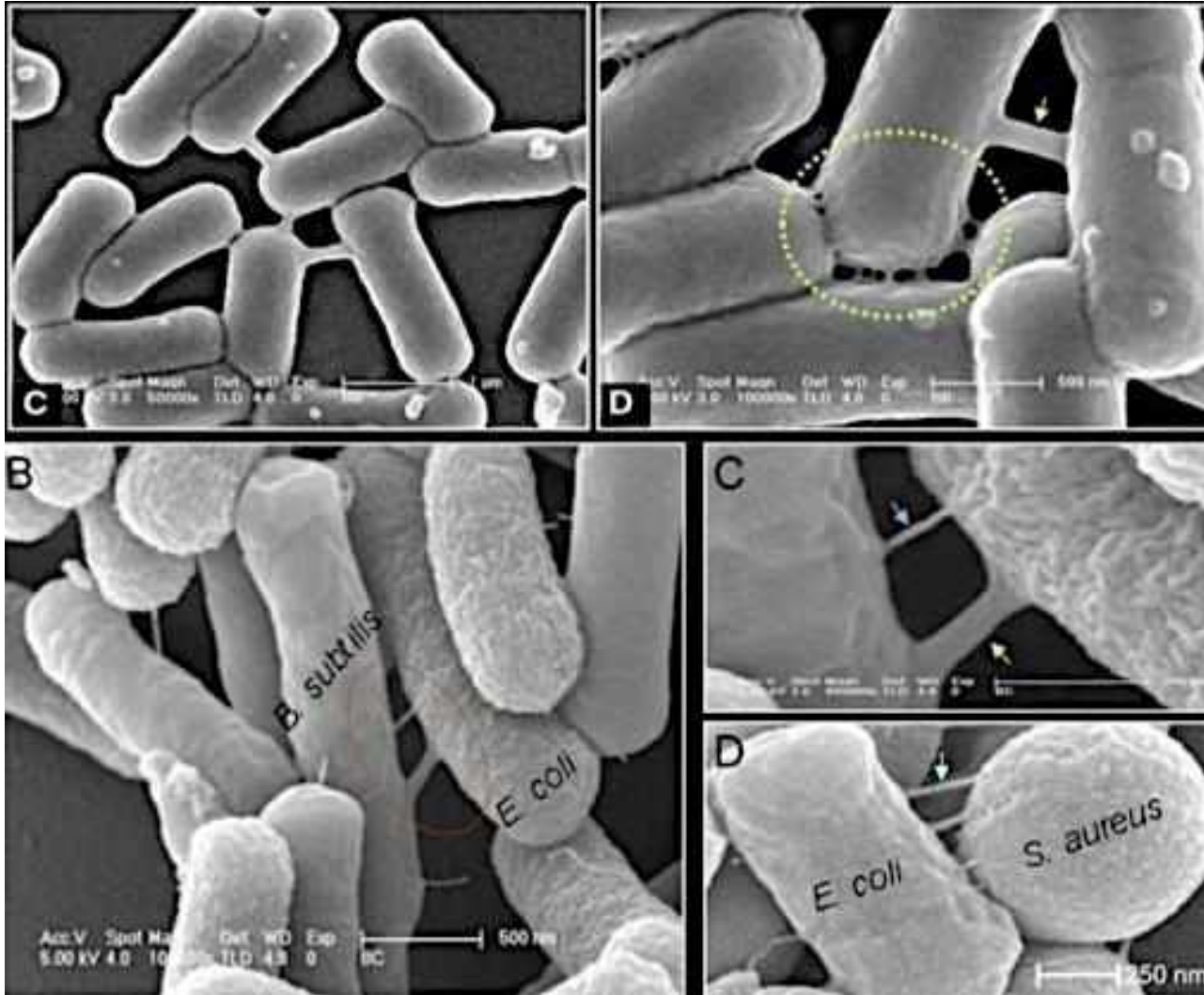
Gene expression

Biofilm provides increased antimicrobial tolerance

Bacteria living in biofilms can be up to 1,000/1500 times more tolerant to antibacterial compounds than their planktonic counterparts



Biofilm production and multidrug resistance (MDR)



Horizontal gene transfer: conjugation, transformation, transduction are increased in biofilms

The biofilm lifestyle also increases plasmid stability and the range of mobile genetic elements

Distinguishing between resistance, tolerance and persistence

Resistance

The genetic ability to counteract antimicrobial treatments.
Resistance is quantified by the minimum inhibitory concentration (MIC)

Tolerance

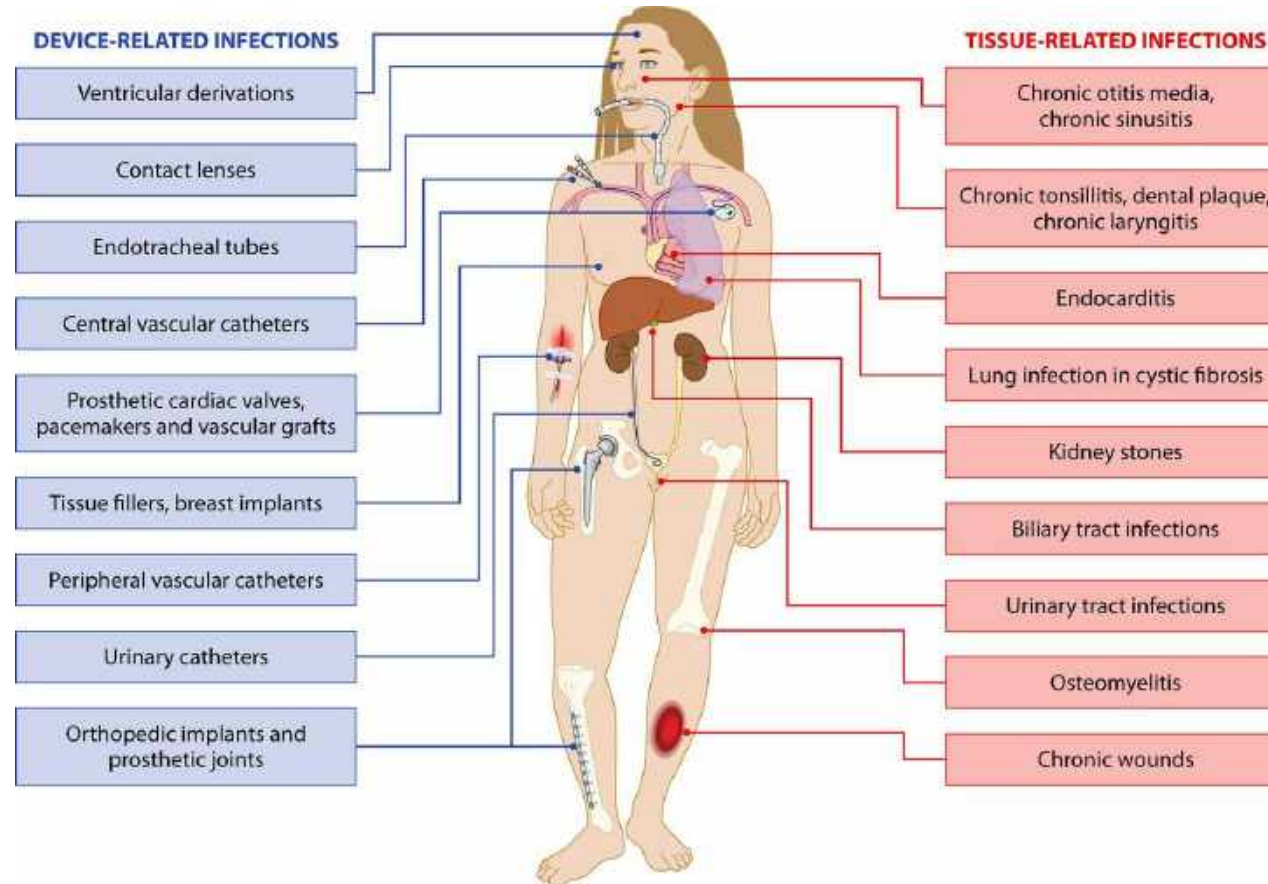
The ability to evade antimicrobial treatments of genetically susceptible microorganisms.
Tolerance can be quantified by the minimum duration for killing (MDK99).

Persistence

Prolongs the duration of treatment that bacteria can sustain only for a subpopulation
(MDK99.99)

Clinical relevance of Biofilm infections

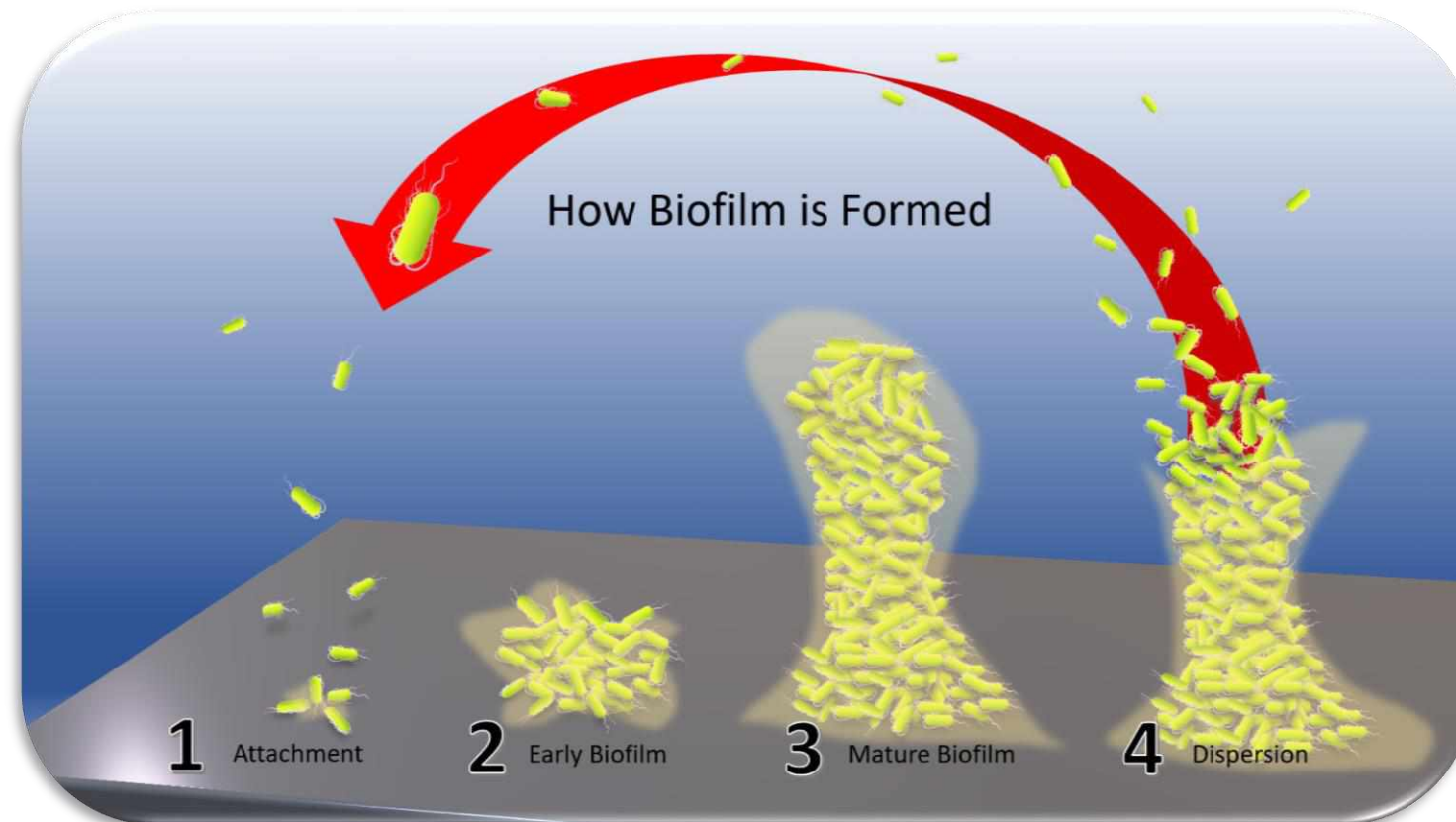
Biofilms are present in more than 80% of all human infections



Biofilms cause over 2 million infections annually, resulting in US\$11B in additional costs

What is the optimal antibiotic strategy for treatment of established biofilm infections?

Antibiograms are performed on planktonic cells and do not take into account biofilm



The opportunity



The Emerging Role of Microbial Biofilm in Lyme Neuroborreliosis

Enea Gino Di Domenico^{1*}, Ilaria Cavallo¹, Valentina Bordignon¹, Giovanna D'Agosto¹, Martina Pontone¹, Elisabetta Trento¹, Maria Teresa Gallo¹, Grazia Prignano¹, Fulvia Pimpinelli¹, Luigi Toma² and Fabrizio Ensoli¹



Article

Biofilm is a Major Virulence Determinant in Bacterial Colonization of Chronic Skin Ulcers Independently from the Multidrug Resistant Phenotype

Enea Gino Di Domenico^{1,2*}, Ilaria Farulla¹, Grazia Prignano¹, Maria Teresa Gallo¹, Matteo Vespaziani¹, Ilaria Cavallo¹, Isabella Sperduti², Martina Pontone¹, Valentina Bordignon¹, Laura Cilli¹, Alessandra De Santis¹, Fabiola Di Salvo¹, Fulvia Pimpinelli¹, Ilaria Lescauni La Pazola², Luigi Toma² and Fabrizio Ensoli¹



Nanometric ion pair complexes of tobramycin forming microparticles for the treatment of *Pseudomonas aeruginosa* infections in cystic fibrosis

Crista Sardo¹, Enea Gino Di Domenico¹, Barbara Pontio¹, Davide De Rocco¹, Roberta Santucci², Fiorentina Ascenzi^{1,3}, Gaetano Giannone⁴, Gemma Cavallaro^{1,5,6}



Development of an *in vitro* Assay, Based on the BioFilm Ring Test[®], for Rapid Profiling of Biofilm-Growing Bacteria

Enea G. Di Domenico^{1*}, Luigi Toma², Christian Provot¹, Fiorentina Ascenzi¹, Isabella Sperduti², Grazia Prignano¹, Maria T. Gallo¹, Fulvia Pimpinelli¹, Valentina Bordignon¹, Thierry Bernardi¹ and Fabrizio Ensoli¹



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RESEARCH ARTICLE

The clinical Biofilm Ring Test: a promising tool for the clinical assessment of biofilm-producing *Candida* species

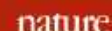
Enea Gino Di Domenico^{1,2,3*}, Ilaria Cavallo¹, Maria Guembe², Grazia Prignano¹, Maria Teresa Gallo¹, Valentina Bordignon¹, Giovanna D'Agosto¹, Isabella Sperduti¹, Luigi Toma² and Fabrizio Ensoli¹



A Novel Approach Based on Low-Field NMR for the Detection of the Pathological Components of Sputum in Cystic Fibrosis Patients

Michela Abenati¹, Fiorentina Ascenzi^{1,2}, Enea Gino Di Domenico^{1,2}, Massimo Marchio³, Alessandra Venturi⁴, Marco Confalonieri⁵, Saide El Gioia⁶, Massimo Cuomo⁶, Barbara Dupas⁶, Gabriele Grassi^{1,6} and Mario Grassi¹

SCIENTIFIC REPORTS



Inflammatory cytokines and biofilm production sustain *Staphylococcus aureus* outgrowth and persistence: a pivotal interplay in the pathogenesis of Atopic Dermatitis

E. G. Di Domenico^{1,2*}, I. Cavallo¹, V. Bordignon¹, G. Prignano¹, I. Sperduti², A. Gurtner³, E. Trento¹, L. Toma², F. Pimpinelli², B. Capitani⁴ & F. Ensoli²



Review

Biofilm Producing *Salmonella* Typhi: Chronic Colonization and Development of Gallbladder Cancer

Enea Gino Di Domenico^{1,2*}, Ilaria Cavallo¹, Martina Pontone¹, Luigi Toma² and Fabrizio Ensoli¹

J. Nanopart. Res. (2016) 18:301
DOI 10.1007/s11051-015-3611-9

RESEARCH PAPER



Positively charged biopolymeric nanoparticles for the inhibition of *Pseudomonas aeruginosa* biofilms

Laura Chronopoulou · Enea Gino Di Domenico · Fiorentina Ascenzi · Cleofe Palocci

BMC Microbiology (2018) 18:103
https://doi.org/10.1186/s12866-018-1103-5

BMC Microbiology

RESEARCH ARTICLE

Open Access

Microbial biofilm correlates with an increased antibiotic tolerance and poor therapeutic outcome in infective endocarditis

Enea Gino Di Domenico^{1*}, Sara Giordano Rimoldi², Ilaria Cavallo¹, Giovanna D'Agosto¹, Elisabetta Trento¹, Giovanni Cagnoni³, Alessandro Palazzi⁴, Cristina Pagani⁴, Francesca Romeri⁵, Erika De Vecchi⁶, Monika Schiavoni⁷, Daniela Secchi⁸, Carlo Antonia⁹, Giuliano Rizzardi¹⁰, Rita Barbara Cicchitto¹¹, Luigi Toma¹², Daniela Kovacs¹³, Giorgio Carlini¹⁴, Maria Teresa Gallo¹, Maria Rita Giandomeni¹⁵ and Fabrizio Ensoli¹



Lucarelli et al. Antimicrobial Resistance and Infection Control (2017) 6:22
DOI 10.1186/s13075-017-0754-4

Antimicrobial Resistance and Infection Control

RESEARCH

Open Access

Ralstonia mannitolilytica infections in an oncologic day ward: description of a cluster among high-risk patients

Cristina Lucarelli^{1,2*}, Enea Gino Di Domenico¹, Luigi Toma², Domenico Iacuzzi³, Grazia Prignano¹, Maria Fortunata¹, Loredana Balaghi⁴, Fabrizio Ensoli¹, Patrizia Picardi⁵, Aurora Garcia-Fernandez⁶, Annalisa Piroli⁷ and Loredana Ingrosso⁸



Review

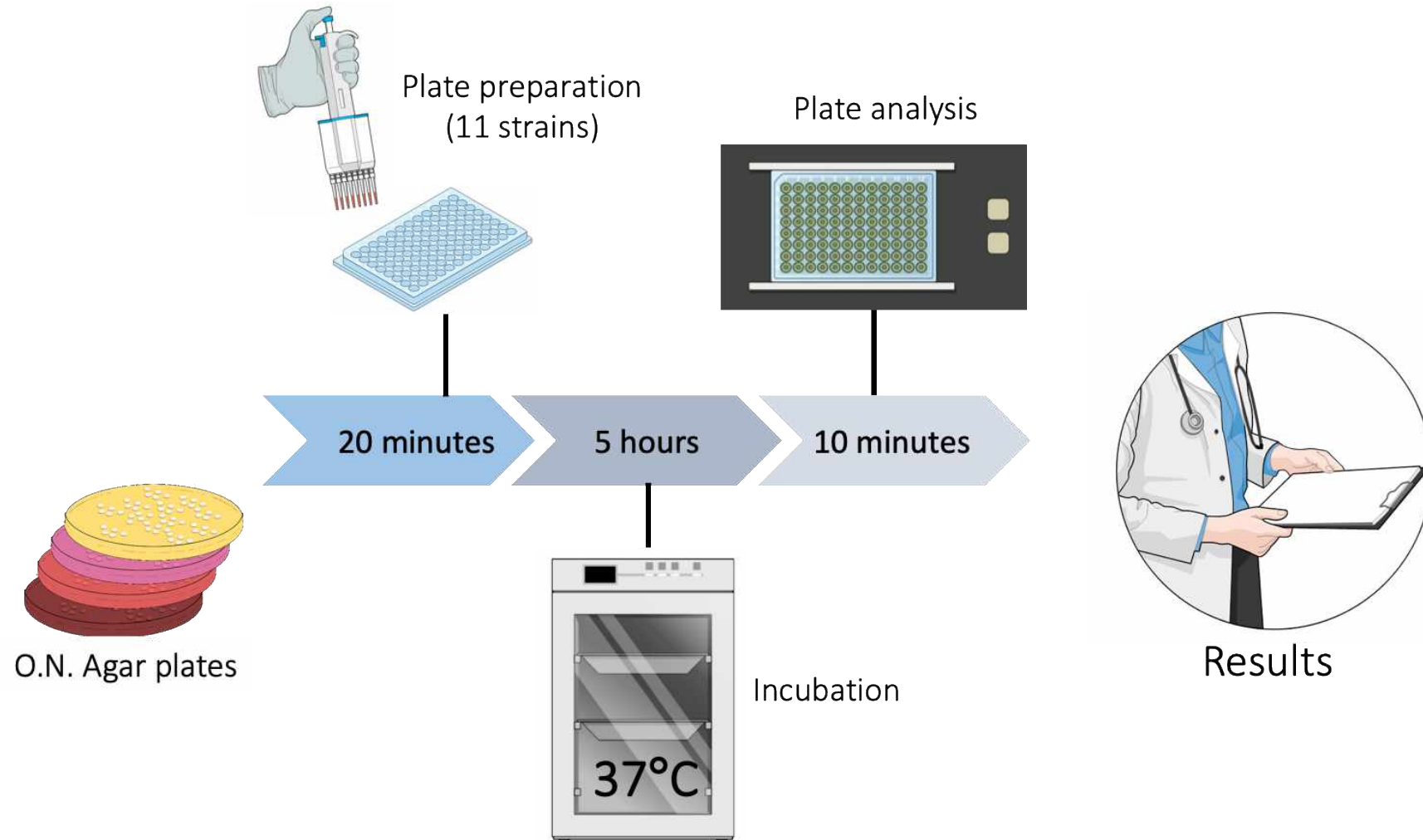
Staphylococcus aureus and the cutaneous microbiota biofilms in the pathogenesis of atopic dermatitis

Enea Gino Di Domenico¹, Ilaria Cavallo¹, Bruno Capitani², Fiorentina Ascenzi¹, Fulvia Pimpinelli¹, Aldo Momonetti³, Fabrizio Ensoli¹

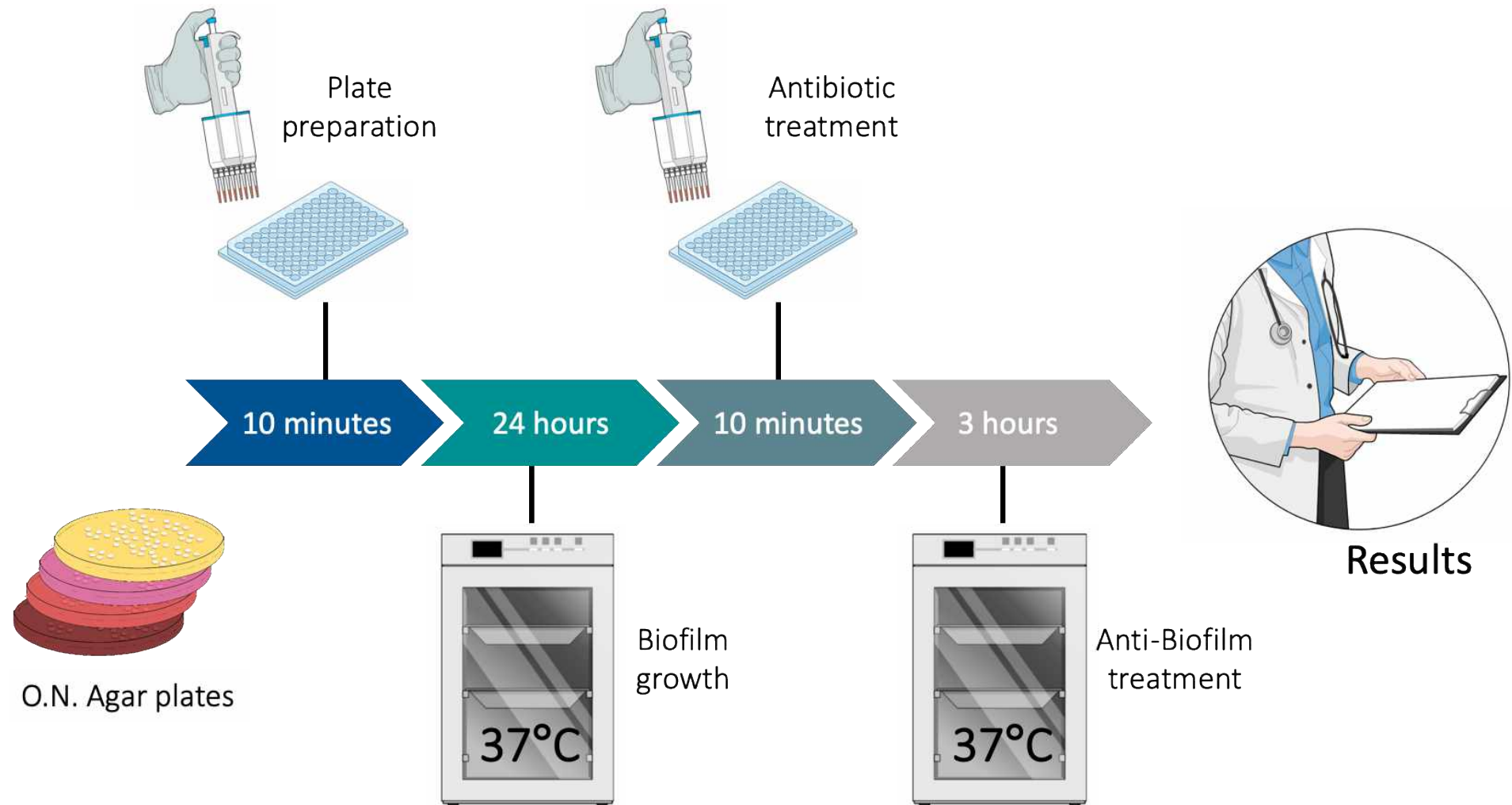


clinical BIOFILM RING TEST[®] (cBRT)

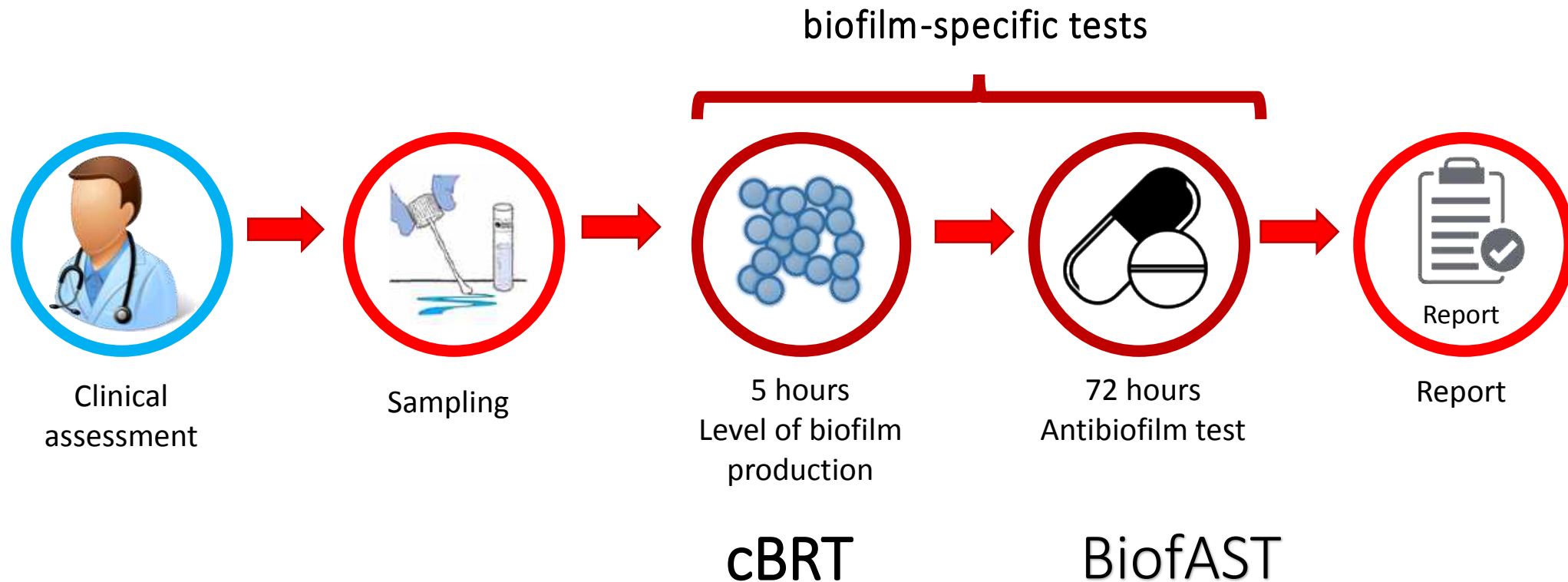
The working protocol



Biofilm Antimicrobial Susceptibility Testing (BiofAST)



Development of innovative strategies for the treatment of biofilm infections



Clinical and microbiological aspects of biofilm-associated surgical site infections



Case presentation

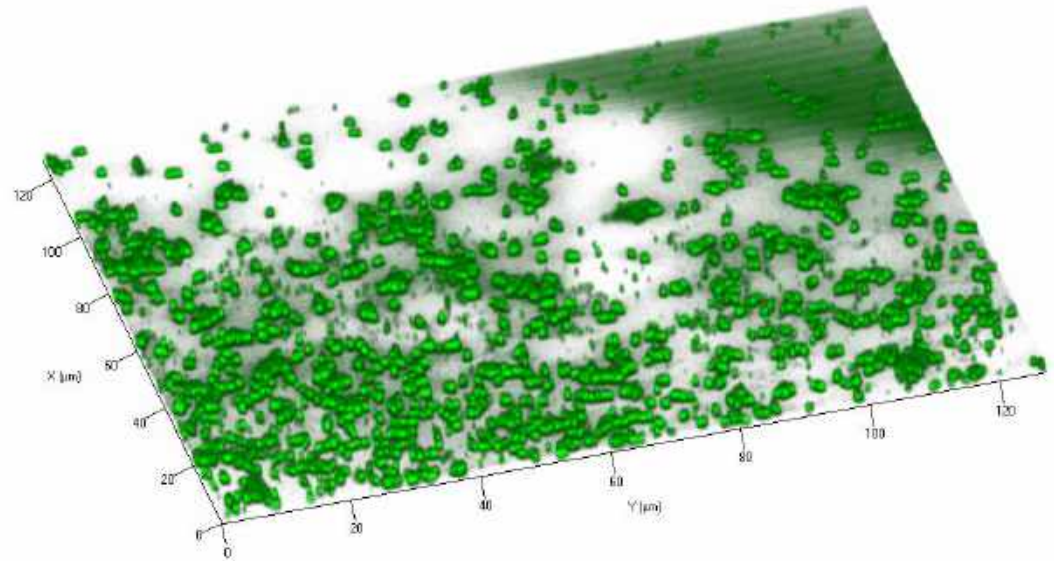
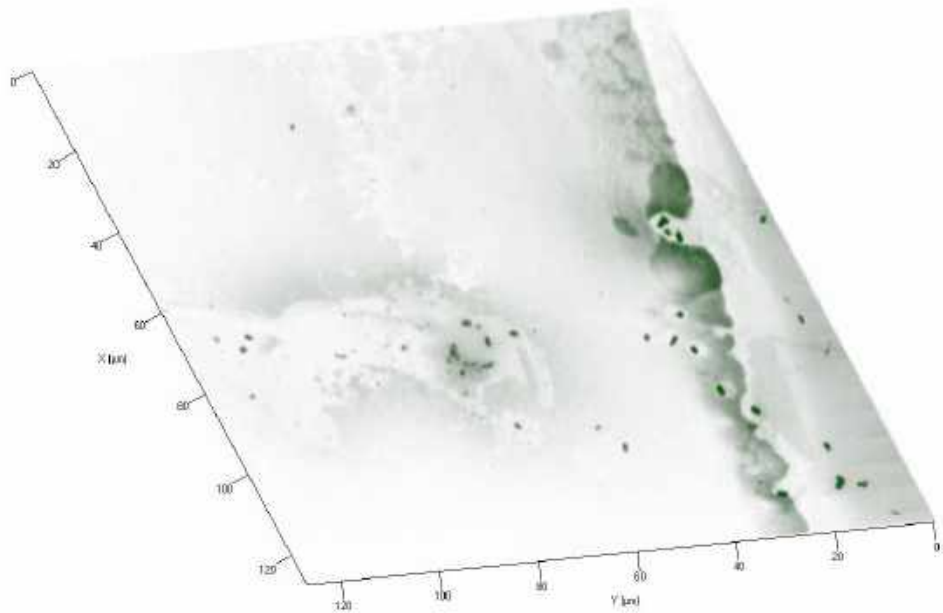
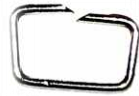
78-Year-old woman with oral cancer

Tracheostomy over the second tracheal ring

Pectoralis major flap for reconstructive head and neck surgery

Surgical site infection by:
Staphylococcus aureus
Pseudomonas aeruginosa

Biofilm on suture



Co-infection by *S. aureus* and *P. aeruginosa*

I.D.: XXX Sig. XXX		Sesso F		MICROBIOLOGIA						
Data di Nascita: XXX		Età: 78 Anni		PROVENIENZA: OTORINO						
				Materiale: TAMPONE ULCERA						
STRAIN:	<i>Staphylococcus aureus</i>				<i>Pseudomonas aeruginosa</i>					
RESULT:	Moderate biofilm producer				High biofilm producer					
	Antimicrobials	MIC (mg/L)	INT	BMIC (mg/L)	INT	Antimicrobials	MIC (mg/L)	INT	BMIC (mg/L)	INT
	Benzilpenicillin	> 0.5	R	> 8	R	Amikacin	≤ 2	S	≤ 2	S
	Clindamycin	≤ 0. 25	S	> 2	R	Cefepime	≤ 1	S	> 32	R
	Daptomycin	≤ 0. 50	S	4	R	Ceftazidime	4	S	32	R
	Erythromycin	> 2	R	4	R	Ciprofloxacin	≤ 0.25	S	> 2	R
	Fusidic Acid	≤ 0,5	S	≤ 1	S	Gentamicin	≤ 1	S	1	S
	Gentamicin	≤ 1	S	1	S	Imipenem	2	I	> 16	R
	Linezolid	2	S	1	S	Meropenem	1	S	> 16	R
	Oxacillin	> 2	R	> 2	R	PIT	8	S	> 128	R
	Rifampicin	-	-	≤ 0.06	S	Piperacillin/Tazobactam (PIT) TMP/SMX = Trimethoprim/Sulfamethoxazole				
	Teicoplanin	≤ 0,5	S	4	R					
	Tigecycline	0.25	S	0.25	S					
	TMP/SMX	≤ 10	S	≤ 10	S					
	Vancomycin	≤ 0,5	S	2	S					

EUCAST Clinical breakpoints - bacteria (v 9.0)



Daptomycin
+
Meropenem



Gentamicin
+
Meropenem

D=0



D=7



D=14



D=21



Conclusions

There are not many answers about how to treat chronic biofilm infections

The strength of biofilm production is a key risk factor affecting the antimicrobial therapy

Early, aggressive and targeted antibiotic treatments against biofilm infections

Biofilm profiling with cBRT and antibiotic efficacy on biofilm-growing microorganisms with BioFAST represent promising tools to predict the clinical outcome of antibiotic therapy

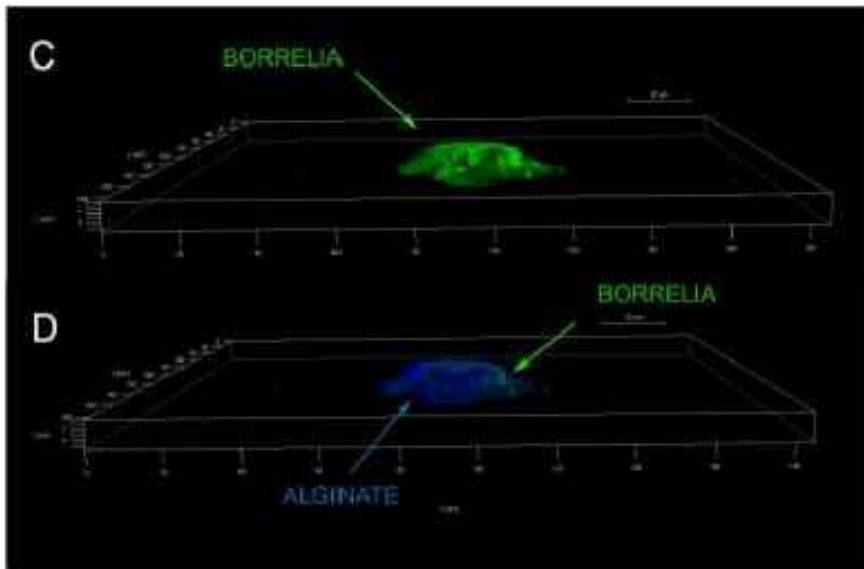
Biofilm Production in *Borrelia burgdorferi*: *in vivo* and *in vitro* evidences

Article

The Long-Term Persistence of *Borrelia burgdorferi* Antigens and DNA in the Tissues of a Patient with Lyme Disease

Eva Sapi^{1,2}, Rumanah S. Kasliwala¹, Hebo Ismail¹, Jason P. Torres¹, Michael Oldakowski¹, Sarah Markland¹, Gauri Gaur¹, Anthony Melillo¹, Klaus Eisendle², Kenneth B. Liegner^{3,4,5}, Jenny Libien⁶ and James E. Goldman⁷

CONFOCAL MICROSCOPY

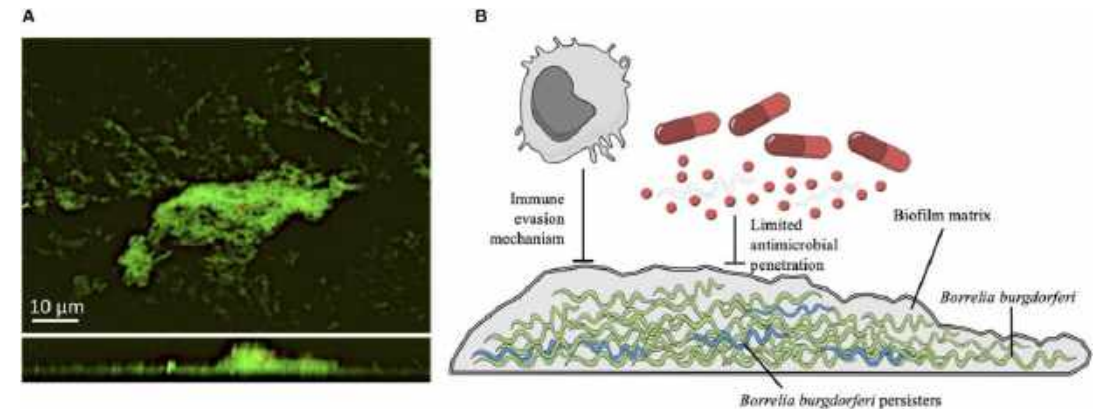


Borrelia biofilm in infected liver tissue



The Emerging Role of Microbial Biofilm in Lyme Neuroborreliosis

Enea Gino Di Domenico^{*}, Ilaria Cavallo¹, Valentina Bordinon¹, Giovanna D'Agosto¹, Martina Pontone¹, Elisabetta Trento¹, Maria Teresa Gallo¹, Grazia Prignano¹, Fulvia Pimpinelli¹, Luigi Toma² and Fabrizio Ensoli¹



Thank you



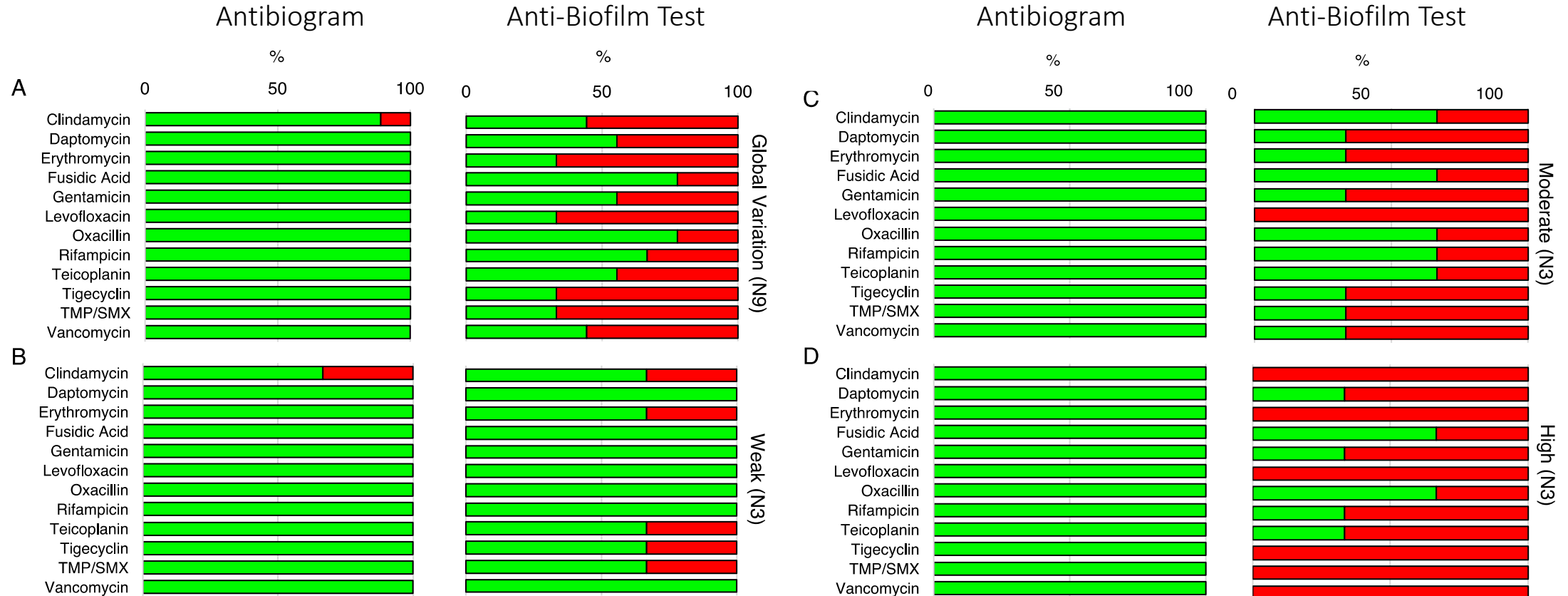
IRE **ISG**
ISTITUTO NAZIONALE TUMORI
REGINA ELENA **SAN GALLICANO**
ISTITUTO DERMATOLOGICO

ISTITUTI DI RICOVERO E CURA A CARATTERE SCIENTIFICO

Microbiology and Virology
San Gallicano Institute
I.F.O. Istituti Fisioterapici Ospitalieri

email: enea.didomenico@ifo.gov.it

Anti-Biofilm Test: the antibiotic profiling on biofilm-producing bacteria



Antibiotic tolerance is linked to the level of biofilm production